

Evaluation of the antimicrobial activity and cytotoxicity of *Rhaponticoides iconiensis* seed extract

Sumeyye OZYAMAN^{1*}, Gamze Nur KOSOVALI², Ibrahim OZTURKCI³, Merve SIMSAR⁴

1 Istanbul Medipol University, Graduate School of Health Sciences, Department of Histology and Embryology, Istanbul, Türkiye

2 Bahcesehir University, School of Medicine, Istanbul, Türkiye

3 MEF University, Computer Science and Engineering, Istanbul, Türkiye

4 Marmara University, Institute of Health Sciences, Istanbul, Türkiye

ABSTRACT

The goal of this study is to evaluate the antimicrobial and cytotoxic activities of aqueous extract from *Rhaponticoides iconiensis* seed. The Soxhlet extraction method was used to extract the seed in distilled water. It was tested for antimicrobial activities against pathogenic bacteria *Escherichia coli* using the disc diffusion method. Additionally, the cytotoxic activity of the seed extract on the MCF7 breast cancer cell line was evaluated by MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) assay. *R. iconiensis* showed strong antibacterial activity against *Escherichia coli* compared to Cefotaxime antimicrobial agents. Additionally, an effective *in vitro* cytotoxic activity against the MCF7 is observed ($p < 0.01$). The present study is the first report of endemic *Rhaponticoides iconiensis* seeds exhibiting potential antimicrobial activity and cytotoxicity, and it requires further investigation and characterization. These findings may be applied as a guideline for selecting Turkish medicinal plant species for further pharmacological and phytochemical studies.

Keywords: *Rhaponticoides iconiensis*, antimicrobial, cytotoxicity, cancer, phytotherapy

*Corresponding author: Sumeyye OZYAMAN

E-mail: sumeyye.ozyaman@medipol.edu.tr

ORCIDs:

Sumeyye OZYAMAN: 0000-0002-6077-7423

Gamze Nur KOSOVALI: 0009-0003-1642-9952

Ibrahim OZTURKCI: 0009-0005-0858-7226

Merve SIMSAR: 0009-0002-8460-5447

(Received 6 Jun 2024, Accepted 21 Jul 2024)

INTRODUCTION

Several plants are regarded as potential candidates for drugs due to their drug-like characteristics¹. Since Türkiye is included in three different flora regions, plant diversity is of great importance. Many plants belonging to these regions are distributed among the plant communities in Türkiye^{2,3}. It is known that there are approximately 9000 different natural plant species in Türkiye and 30% of these plants are endemic. Although there are many endemic plants, this diversity of plants cannot be utilized sufficiently⁴.

Cancer is the primary cause of mortality in every nation worldwide⁵. Based on 2019 estimates, the World Health Organization (WHO) indicated that in 112 of 183 countries, cancer was the first or second most common cause of death before the age of 70, and in 23 countries, it was the third or fourth leading cause⁶. Although there is a significant decrease in stroke and coronary heart disease death rates compared to cancer, this is still the leading cause of death⁷. GLOBOCAN 2020 estimated that cancer claimed 10 million lives with 19.3 million new cases diagnosed in 2020⁸.

Asteraceae is a vast and globally distributed family of flowering plants, with over 1,100 genera and 2,500 species. Within this family, *Centaurea L.* genus is among the largest and the most significant genera within the Asteraceae family and the *Rhaponticoides* genus is a member of the Asteraceae family. *Rhaponticoides iconiensis* (*R. iconiensis*), a species within this genus, is endemic to Konya, Türkiye⁹. It is widely used as herbal medicine due to its various properties⁶. In Türkiye, there are 8 species belonging to the *Rhaponticoides* genus with 7 of them being endemic. These species are known by the Turkish name "Tülüşah"^{4,10}. In our study, we focused on *R. iconiensis* which is one of these species.

There are few studies that reported chemical composition and bioactivity of *R. iconiensis*. Paşayeva et al. showed the antioxidant and antidiabetic activity of *R. iconiensis* flower¹¹. Additionally, antioxidant properties, total phenol amounts and flavonoid amounts of the endemic *Rhaponticoides* species were determined. The mean percentage of DPPH (2,2-diphenyl-1-picrylhydrazyl) antioxidant free radical scavenging effect values is presented¹². The antioxidant properties, enzyme inhibition, and levels of phenolics and flavonoids in methanol extracts (obtained via maceration and Soxhlet extraction) and extracts of water (prepared through infusion) were assessed of leaves, roots, and flower heads of *R. iconiensis*. It was reported that the leaf extracts of *R. iconiensis* had higher levels of phenolic and flavonoid compounds compared to

the flower and root extracts. Also, the extract had 87 compounds, including hydroxybenzoic, hydroxycinnamic, and acylquinic acids, anthocyanins, flavones and flavanones. On the other hand, it has been indicated that *R. iconiensis* might be a valuable source of natural enzyme inhibitors for developing new drugs to address global health issues because of its enzyme inhibitory effects¹³.

Despite all aforementioned data, there is no study evaluated effect of *R. iconiensis* seed extract. We aimed to uncover the antimicrobial activity and cytotoxicity of aqueous *R. iconiensis* seed extract.

METHODOLOGY

Plant material

The endemic *R. iconiensis* (Hub.-Mor.) M.V. Agab. & Greuter (Voucher No. 11.048) used in our studies was obtained from the Nezahat Gökyiğit Botanical Garden with its species identification confirmed. A very small population of *R. iconiensis* was found between the canal and the highway near the village of Orta Karaören, 18 km from Seydisehir, by the expert of the subject Prof. Dr. Mecit Vural.

Solvent extraction of plants

R. iconiensis seeds were weighed to 10 g in a Soxhlet extractor thimble and placed in the extraction apparatus. Seeds were extracted in a 250 ml conical flask based on the feed-to-solvent ratio [1:10 (w/v)]. The extraction was performed on 10 g of seed using 100 ml of water. A heating mantle was utilized to reflux the mixture for extraction periods spanning from 6 to 8 hours¹³. Once the extraction time was completed, the extract solution was cooled at room temperature. It was subsequently filtered using cone filter paper (Whatman no.1) and left in a water bath¹⁴. Powder sample was measured and stored at -4°C for further analysis.

Cell culture and cell line

For the assessment of the anticancer activity of the seeds, MCF7 human breast cancer cell line was used. L929 mouse fibroblast were used as a negative control in the study. MCF7 and L929 cells were cultured in Dulbecco's Modified Eagle Medium/Nutrient Mixture (DMEM) medium supplemented with 10% FBS (Fetal Bovine Serum), 1% penicillin-streptomycin (P4458; Sigma, USA) and 1% L-glutamine at 37°C with 5% CO₂ in a humidified incubator. Cells were suspended in medium with 1% penicillin-streptomycin (P4458; Sigma, USA) and 1% L-glutamine. Then, cells were counted (at a density of 2×10⁴ cells/ml), transferred into a 96-well plate, and incubated for 24 hours prior to the addi-

tion of extract. The seed extracts were dissolved in cell culture medium, then diluted to different serial concentrations. The cells were treated with 10, 50, 100 µg/ml dilutions of the seed extracts and 1% Triton-x was added as a positive control. Untreated cells given the same volume of medium considered as the control.

Cell viability assay

For cell viability assay, it was measured using the standard colorimetric MTT (3-(4, 5-dimethylthiazol-2-yl)-2, 5-diphenyl tetrazolium bromide) assay technique. After 24 hours incubation, 10 µL of MTT solution (5 mg/ml) was dispensed into each well and incubated for 4 hours. Subsequently, 100 µL of DMSO was added to each well to dissolve the formazan crystals. Then absorbance readings were recorded at 570 nm using a microplate reader for each cell line. The cytotoxicity was determined by comparing the absorbance levels between the serial dilutions and the control samples¹⁵.

Antimicrobial assay

Escherichia coli (*E. coli*) (ATCC 25922) was obtained from the Department of Clinical Microbiology, Faculty of Medicine at Istanbul University. The organisms were kept in Nutrient agar at 4°C until analyses.

Disc diffusion method

The antimicrobial effect of *R. iconiensis* was tested according to the disc diffusion Kirby-Bauer technique, which complies with the standards recommended by CLSI, 2015¹⁶. To prepare the bacterial inoculum, the bacterial strain was suspended in sterile dH₂O, and the turbidity was adjusted to approximately 10⁶ CFU/ml, equivalent to 0.5 McFarland standards¹⁷. Nutrient agar (4018102; Biolife, Italy) was used for antimicrobial activity. Next, 28 g of agar was dissolved in 1 liter of dH₂O, autoclaved and allowed to cool. Then, 20 ml of agar was poured into petri dishes. The sterile swab was immersed in the standardized bacterial suspension and utilized to uniformly inoculate the bacteria onto Nutrient agar plates. The plates were left to air dry for 5 minutes and Whatman no.1 filter paper was utilized to produce discs with a diameter of 6 mm. These discs were sterilized via autoclaving and subsequently dried in a hot air oven at 80°C for one hour. Circular discs measuring 6 mm in diameter were created by punching through a sheet of filter paper. Each disc was impregnated with 20 µl each of 1 g/ml *R. iconiensis* aqueous extract, Cefotaxime antimicrobial agents (dissolved in dH₂O) as a positive control, or dH₂O alone as a negative control. Following impregnation, all papers were dried, and the discs were maintained under sterile conditions until further use. Then, all discs were positioned on

the plates with flamed forceps and delicately pressed to guarantee complete contact with the agar. The plates were subsequently placed in an incubator at 37°C for a period of 24 hours. After incubation, the areas surrounding the discs where bacterial growth was impeded were measured and recorded in millimeters. The experiments were replicated six times to ensure consistency and reliability.

Statistical analysis

Statistical evaluations of the results were performed using the ANOVA test with GraphPad InStat (GraphPad Software Inc., San Diego, CA, USA). Tests were conducted with a 95% confidence interval, and $p < 0.05$ was considered significant. Data averages for the groups are presented as means \pm standard deviation (SD). To determine the concentration necessary for a 50% reduction in cell viability (IC_{50}), regression analysis was used, and graphs were prepared using Microsoft Office Excel 2017.

RESULTS and DISCUSSION

Many studies show that plants have a potential for use in traditional medicines in numerous countries worldwide. Moreover, plants are identified as a crucial source for discovering new cytotoxic compounds, with many polyphenolic flavonoids having antitumor effects¹⁸. Every part of the plant, including the leaf, flower, seed, and rhizome, is known to be used for both nutritional and medicinal purposes¹⁹. Extracts from plant flowers, fruits, and seeds have shown a variety of biological activities in numerous studies^{20,21}. Despite the limited research has been conducted on endemic *R. iconiensis* to investigate its biological activities with some parts of it, there is an absence of studies regarding the activity of the seed. Herein, we document the cytotoxic and antimicrobial effects of seed samples from the endemic *R. iconiensis* species to elucidate their biological activities.

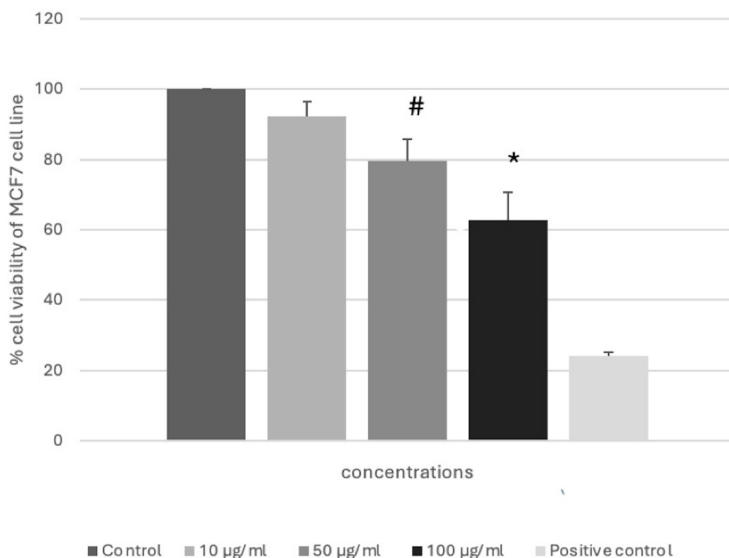
Cytotoxic activity findings

In our study, MTT test was performed to reveal the cytotoxic effects of the aqueous seed of endemic *R. iconiensis* species on the MCF7 breast cancer cell line.

Breast cancer is the leading cause of death and the most prevalent type of cancer among women globally, with over 10 million new cases identified annually. The disease's progression, prognosis, and treatment outcomes are influenced by its heterogeneity. According to the WHO, breast cancer has the highest prevalence worldwide. In 2018, breast cancer caused to 2.09 million diagnoses

and 627,000 deaths among women. Moreover, around 1.7 million new cases were reported in 2020, resulting in an estimated 627,000 deaths²². Therefore, we showed that *R. iconiense* could be a new potential candidate for MCF7 breast cancer cell line.

In a study, it is investigated the methanol extracts and sub-extracts derived from the leafy stems and flowers of *R. iconiense*. Their cytotoxic effects were examined on A549, Colo 205, HepG2, Beas-2b and MCF7 cell lines using the MTT assay. The constituents of *R. iconiense* extracts were identified and quantified using LC–MS/MS. Consequently, while the methanol extracts had no cytotoxicity against A549 cells, they showed cytotoxic effects on HepG2, Colo 205, MCF-7, and Beas-2b cell lines²³. To determine whether seed of it has anti-cancer effect, we first explored the cytotoxicity of aqueous *R. iconiense* *in vitro* and we showed the decreased % viability rate of the MCF7 cell line, and the results were statistically significant with increasing dose (Figure 1).



* $p < 0.01$ compared to the control group

Figure 1. Representing the percentage (%) cell viability of *R. iconiense* on MCF7 cell line. $p < 0.05$. Values in the groups are expressed as mean (X) \pm standard deviation (SD).

To investigate the cytotoxic effects of *R. iconiensis*, it was tested on healthy cell lines. After treating the L929 mouse fibroblast cells with different concentration of seed extract, there was no significant difference in viability of L929 compared to control (Figure 2).

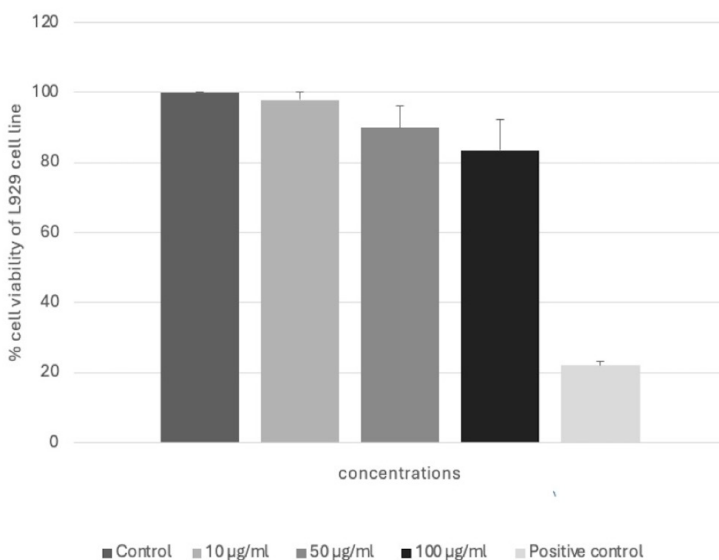


Figure 2. Representing the percentage (%) cell viability of *R. iconiensis* on L929 cell line. Results compared to control group. Values in the groups are expressed as mean (X) ± standard deviation (SD).

The IC_{50} (the dose that inhibits cell growth by 50%) value of *R. iconiensis* was found 56.33 µg/ml in MCF7 cell line. Tugay et al. provided that sub-extracts of leafy stem and flowers of *R. iconiensis* were most effective against MCF7 cancer cell line²³. In a study, the cytotoxic effects of methanol extracts from the stem and flowers of *R. iconiensis* were showed against the A-549, HEPG2, MCF7, COLO-205, and BEAS-2b cell lines by MTT and SRB methods²⁴. Hence, in present study we have proven that seed extracts of *R. iconiensis* have cytotoxic effect on MCF7 cell line.

Consequently, *R. iconiensis* showed cytotoxic effect on MCF7 cell line depending on increasing doses [50 µg/ml ($p < 0.05$) and 100 µg/ml ($p < 0.01$)] while it has not showed reduction in cell viability on L929 mouse fibroblast cell line.

Antimicrobial activity findings

The type of *E. coli* used in molecular biology labs is a model commensal bacterium that is prevalent in the mammalian intestine. However, some strains of this species can cause serious illnesses in humans²⁵. Given this, the purpose of the present study was to determine the antimicrobial activity of *R. iconiensis* against *E. coli* and its antimicrobial activity has been demonstrated compared to Cefotaxime antimicrobial agents (Table 1). Paşayeva et al. reported the similar positive results, and they showed the antimicrobial effect with the microdilution method using leafy stem extracts of *R. iconiensis*²⁶.

Table 1. Zones of inhibitions as shown by aqueous *R. iconiensis* seed extract against *E. coli*. Data represents Mean \pm Standard error of mean (n=6).

Disc diffusion zone diameters (cm) \pm SD			
Microorganism	Positive control (Cefotaxime)	<i>R. iconiensis</i>	Negative control (dH ₂ O)
<i>E. coli</i>	4.9 \pm 0,25	3,2 \pm 0,2	—

*(— = no zone)

In summary, we have described robust antimicrobial activity and cytotoxic effect of seed samples from the endemic *R. iconiensis* which is extracted in distilled water by Soxhlet extraction method, we used only aqueous extract and focused exclusively on a single cell line and one bacterial strain. Hence, additional research encompassing a wider variety of cell lines and bacterial strains will provide a more thorough comprehension of their therapeutic potential.

STATEMENT OF ETHICS

No need for ethical approval for this study.

CONFLICT OF INTEREST STATEMENT

The authors state that they have no conflicts of interest.

AUTHOR CONTRIBUTIONS

All authors contributed to the work equally.

FUNDING SOURCES

No financial support was received for the study.

ACKNOWLEDGMENTS

This research was funded by the Bayrampasa Science Center.

REFERENCES

1. Bernhoft A. A brief review on bioactive compounds in plants. In: Norwegian Academy of Science and Letters. Bioactive compounds in plants - benefits and risks for man and animals. Proceedings of a symposium; 2008, Nov 13-14; Oslo, Norway. Norwegian Academy of Science and Letters; 2010. p. 11-17.
2. Svenning JC, Skov F. The relative roles of environment and history as controls of tree species composition and richness in Europe. *J Biogeogr*, 2005;32:1019-1033. Doi: 10.1111/j.1365-2699.2005.01219.x
3. Avcı M. Diversity and endemism in Turkey's vegetation. *J Geography-Cografya Dergisi*, 2005;13:27-55.
4. Güner A, Aslan S. List of plants of Turkey: (vascular plants). Nezahat Gökyiğit Botanical Garden Publications; 2012.
5. Bray F, Laversanne M, Weiderpass E, Soerjomataram I. The ever-increasing importance of cancer as a leading cause of premature death worldwide. *Cancer*, 2021;127:3029-3030. Doi: 10.1002/cncr.33587
6. World Health Organization. Global health estimates 2020: deaths by cause, age, sex, by country and by region, 2000–2019 [Internet]. Geneva: World Health Organization; 2020 [cited 6 Jun 2024]. Available from: <https://www.who.int/data/gho/data/themes/mortality-and-global-health-estimates/ghe-leading-causes-of-death>
7. Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J for Clin*, 2021;209-249. Doi: 10.3322/caac.21660
8. Anderberg A, Baldwin B, Bayer R, Breitwieser J, Jeffrey C, Dillon M, et al. The families and genera of vascular plants. In: Kubitzki K, editor. The families and genera of vascular plants. Vol. 8, Flowering plants. Eudicots. Asterales. Berlin: Springer-Verlag; 2007. p. 561-568.
9. Köse YB, İşcan G, Göger F, Akalın G, Demirci B, Başer KHC. Chemical composition and biological activity of *Centaurea baseri*: new species from Turkey. *Chem Biodivers*, 2016;13:1369-1379. Doi: 10.1002/cbdv.201600070
10. Cinbilgel I, Eren Ö, Duman H. *Rhaponticoides gokceoglu* (Asteraceae), a striking new species from Turkey. *Phytotaxa*, 2014;170(2). Doi: 10.11646/phytotaxa.170.2.5
11. Paşayeva L, Fatullayev H, Celik I, Unal G, Bozkurt NM, Tugay O, et al. Evaluation of the chemical composition, antioxidant and antidiabetic activity of *Rhaponticoides iconiensis* flowers: effects on key enzymes linked to type 2 diabetes *in vitro*, *in silico* and on alloxan-induced diabetic rats *in vivo*. *Antioxidants*, 2022;11:2284. Doi: 10.3390/antiox11112284
12. Hameed YAH, Bağcı Y. Biochemical studies on two endemic *Rhaponticoides* species (*R. aytachii*, *R. iconiensis*). *Eur J Sci Educ*, 2020;417-422.
13. Zheleva-Dimitrova D, Zengin G, Sinan KI, Yıldızıtugay E, Mahomoodally MF, Ak G, et al. Identification of bioactive compounds from *Rhaponticoides iconiensis* extracts and their bioactivities: an endemic plant to Turkey flora. *J Pharm Biomed Anal*, 2020;190:113537. Doi: 10.1016/j.jpba.2020.113537
14. Alara OR, Abdurahman NH, Ukaegbu CI. Soxhlet extraction of phenolic compounds from *Vernonia cinerea* leaves and its antioxidant activity. *J Appl Res Med Aromat Plants*, 2018;11:12-17. Doi: 10.1016/j.jarmap.2018.07.003

15. Mosmann T. Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. *J Immunol Methods*, 1983;65:55-63. Doi: 10.1016/0022-1759(83)90303-4
16. Clinical and Laboratory Standards Institute (CLSI). *Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically*, Approved Standard, 10th ed. National Committee for Clinical Laboratory Standards, Wayne, USA, 2015; p. 88.
17. Cockerill FR, Wikler MA, Alder J, Dudley MN, Eliopoulos GM, Ferraro MJ, et al. *Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically*. Approved Standard, 9th ed. Wayne; 2012.
18. Sak K. Characteristic features of cytotoxic activity of flavonoids on human cervical cancer cells. *Asian Pac J Cancer Prev*, 2014;15(19):8007-8019. Doi: 10.7314/apjcp.2014.15.19.8007
19. Sahu B, Sahu M, Sahu M, Yadav M, Sahu R, Sahu C. An updated review on *Nelumbo nucifera* Gaertn: chemical composition, nutritional value and pharmacological activities. *Chem Biodivers*, 2024;21(5):e202301493. Doi: 10.1002/cbdv.202301493
20. Keskin C. Antioxidant, anticancer and anticholinesterase activities of flower, fruit and seed extracts of *Hypericum amblysepalum* HOCHST. *Asian Pac J Cancer Prev*, 2015;16(7):2763-2769. Doi: 10.7314/apjcp.2015.16.7.2763
21. Zhang X, Bai Y, Wang Y, Wang C, Fu J, Gao L, et al. Anticancer properties of different solvent extracts of *Cucumis melo* L. seeds and whole fruit and their metabolite profiling using HPLC and GC-MS. *Biomed Res Int*, 2020;5282949. Doi: 10.1155/2020/5282949
22. World Health Organization. Accessed 6 June 2024. <https://www.who.int/news-room/fact-sheets/detail/breast-cancer>
23. Tugay O, Paşayeva L, Demirpolat E, Şahin M. Comparative evaluation of cytotoxicity and phytochemical composition of *Centaurea iconiensis* (*Rhaponticoides iconiensis*). *Iran J Sci Technol Trans A: Sci*, 2021;45:65-75. Doi: 10.1007/s40995-020-01030-y
24. Demirpolat E, Paşayeva L, Tugay O. Comparative evaluation of the cytotoxic effects of stem and flower extracts of *Rhaponticoides iconiensis* (Hub.-Mor.) M.V.Agab. & Greuter. *Proceedings*, 2017;1(10):1057. Doi: 10.3390/proceedings1101057
25. Kaper JB, Nataro JP, Mobley HL. Pathogenic *Escherichia coli*. *Nat Rev Microbiol*, 2004;2(2):123-140. Doi: 10.1038/nrmicro818
26. Paşayeva L, Ince U, Fatullayev H, Ceylan C, Tugay O. Hypoglycemic, antioxidant, antimicrobial activities and phytochemical analyses of *Rhaponticoides iconiensis* leafy stem extracts. *Vegetos*, 2021;34:592-599. Doi: 10.1007/s42535-021-00250-w