

Essential oil composition of *Isatis floribunda* Boiss. ex. Bornm. and acetylcholinesterase inhibitory activity of its extract

Elif GÜZEL^{1,2*}, Yasemin Yücel YÜCEL^{3,4}, Mustafa ALKAN⁵, Kaan POLATOĞLU⁶

1 Istanbul Medipol University, Institute of Health Sciences, Istanbul, Türkiye

2 Istanbul Medipol University, School of Medicine, Department of Medical Pharmacology, Istanbul, Türkiye

3 Altinbas University, Faculty of Pharmacy, Department of Biochemistry, Istanbul, Türkiye

4 Altinbas University, Natural Products Research and Development Center (DÜAGEM), Istanbul, Türkiye

5 Yozgat Bozok University, Faculty of Agriculture, Department of Plant Protection, Yozgat, Türkiye

6 Altinbas University, Faculty of Pharmacy, Department of Pharmacognosy, Istanbul, Türkiye

ABSTRACT

This study aims to investigate the essential oil composition of aerial parts of *Isatis floribunda*, and the acetylcholinesterase inhibitory activity of its extracts. Essential oil of the aerial parts of the plant material was obtained and the GC-MS analysis was performed. Then, extracts of the plant material were obtained, and *in vitro* acetylcholinesterase inhibitory assay was performed. GC-MS analysis demonstrated that the composition of *Isatis floribunda* is rich in fatty acid compounds, with the major compounds including dodecanoic acid, nanocosane, hexadecenoic acid, tetradecanoic acid, methyl octadecenoate, decanoic acid, and hexahydrofarnesyl acetone. Methanolic extract of the plant species has demonstrated strong acetylcholinesterase inhibitory activity with the IC₅₀ value of 0.16 mg/mL. Essential oil composition of *Isatis floribunda* has been determined to be rich in fatty acid components, these compounds could demonstrate potent acetylcholinesterase inhibition. Therefore, a novel medication from *Isatis floribunda* extracts could be discovered against Alzheimer's disease.

Keywords: *Isatis floribunda*, acetylcholinesterase inhibition, Brassicaceae, GC-MS

*Corresponding author: Elif GÜZEL

E-mail: elif.guzel@medipol.edu.tr

ORCID:

Elif GÜZEL: 0000-0002-2737-0568

Yasemin Yücel YÜCEL: 0000-0002-6595-6850

Mustafa ALKAN: 0000-0001-7125-2270

Kaan POLATOĞLU: 0000-0001-5493-2995

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INTRODUCTION

Natural sources have been used for developing medicinal agents for centuries. Many modern medicines today have been developed from natural sources such as plants. Traditional medicine that suggests using plant extracts and essential oils of plants still provides treatment to most of the world's population¹. Brassicaceae (Cruciferea) family is a major family which includes approximately 350 genera and 3000 species and distributes mainly in North Temperate Zone. The family species of Brassicaceae, also known as mustards, have been used for centuries for food and herbal remedy purposes². Plants in the Brassicaceae family have been used for their antifungal, antibacterial, antidiabetic and anticancer properties³. Genus *Isatis* is one of the the most extensive genera in the Brassicaceae family, and they are known as biennial, herbaceous shrubs with yellow flowers. Leaves of the *Isatis* genera are known to have antiviral, antibacterial, anticancer, astringent, and febrifuge activities. *Isatis floribunda* is a member of the Brassicaceae family, and an endemic plant distributed in Türkiye and Iran. Mediterranean regions are the major distribution zones of *Isatis floribunda*, including Adana, Çankırı, Ankara, Eskişehir, Kayseri, Konya, and Nevşehir provinces⁴.

In previous studies, extracts of *Isatis floribunda* have been analyzed for their antioxidative, antibacterial, and cytotoxic properties. Plant extracts have demonstrated rich phenolic and flavonoid content, which results in antioxidative activity. Additionally, extracts of *Isatis floribunda* demonstrated antibacterial activity against various Gram-positive and Gram-negative bacteria. Furthermore, HPLC analysis of the extracts of *Isatis floribunda* has shown to be rich in chlorogenic acid, quercetin, p-coumaric acid and caffeic acid^{4,5}. Overall, *Isatis floribunda* extracts have been used for their various biological activities for decades. However, the possible biological activities of *Isatis floribunda* are still yet to be determined.

Alzheimer's disease is the most common type of dementia worldwide. It is characterized by dementia; however, the disease's pathogenesis is still could not be elucidated. The cholinergic hypothesis is believed to be the reason for the disease formation and progression. The cholinergic hypothesis is explained by insufficient endogenous acetylcholine in the cholinergic system⁶. Acetylcholine is a neurotransmitter that causes the signal transmission from one cholinergic neuron to another. Acetylcholine deficiency causes impaired memory, leading to dementia symptoms⁷. One reason for acetylcholine deficiency is the increased acetylcholinesterase (AChE) enzyme activity. AChE enzyme is responsible for the degradation of acetylcholine in the synaptic junction. Overactivation of the AChE enzyme can cause fast degradation of acetylcholine, therefore, deficiency

of acetylcholine, which leads to dementia symptoms⁸. Another hypothesis includes inhibition of the choline-acetyltransferase enzyme, which is the enzyme responsible for the synthesis of acetylcholine. Therefore, an insufficient amount of acetylcholine causes dementia symptoms. Depending on the first hypothesis, AChE inhibitors are used in the treatment of Alzheimer's disease, including Galantamine, Rivastigmine, Physostigmine, Tacrine, and Donepezil^{9,10}. AChE inhibition is the essential treatment strategy for Alzheimer's disease. Therefore, emerging novel AChE inhibitors are essential treatment options¹¹.

In this study, the essential oil composition of *Isatis floribunda* was analyzed with GC-MS analysis, and the AChE inhibitory activity of the extracts of *Isatis floribunda* was tested to determine if the plant species is a promising compound in developing novel treatment options for Alzheimer's disease.

METHODOLOGY

Plant material

Isatis floribunda was collected from its natural habitat during the flowering season from Beypazarı, Ankara, Türkiye. The plant material was deposited to the Gaziosmanpaşa University Herbarium with the voucher specimen number GOPU 3028. Aerial parts of *Isatis floribunda* were used to obtain essential oil and extracts.

Isolation of essential oil

100 grams of dried aerial parts of the plant material were used to obtain the essential oil. Hydro-distillation method was used by Clevenger-type apparatus for 3 hours. At hour 3, the yield of the essential oil was 0.03%.

GC-MS analysis of essential oil

The essential oil analysis was conducted by Agilent 5977 MSD GC-MS system operating in EI mode. Temperatures of the MS transfer line and injector were adjusted at 250 °C, and splitless injection was used through the analysis. As a carrier gas, helium was used with a constant flow rate of 1 mL/min. Innowax FSC column (60 m x 0.25 mm, 0.25 µm film thickness) was used, and the temperature was adjusted at 60 °C for 10 minutes and increased to 220 °C at the rate of 4 °C/min. The temperature was kept stable at 220 °C for 10 min and then increased to 240 °C at a rate of 1 °C/min. Mass spectra were recorded at 70 eV with the mass range m/z 35 to 425. The relative percentage amounts of the separated compounds were calculated from the integration of the peaks in mass spectrum chromatograms. The results of the GC-MS analysis are provided in "Table 1".

Preparation of plant extracts

Air-dried plant material was ground to obtain a fine powder. Then, 10 grams of plant species were used to obtain dichloromethane and methanol extracts of the plant species by the Soxhlet extraction method for 3 hours, respectively. Finally, the excess solvent was evaporated with a rotary evaporator (Heidolph, Germany). The yields of the dichloromethane and methanol extracts were calculated as 5.89% and 34.73%, respectively.

In vitro acetylcholinesterase inhibition assay

The previously described method was used for the AChE inhibition assay¹². The solution of the AChE inhibitory assay contained 240 μL , 1.25 mM 5,5-dithio-bis-(2-nitrobenzoic acid) (DTNB), 192 μL acetylthiocholine iodide (AChI), 1200 μL , 100 mM Tris-HCl buffer pH 8.0 and 20, 40 and 60 μL extract solution. The inhibitory activity was tested for 10 mg/mL concentration stock solution in methanol. In addition, 3 different volumes were tested, including 20 μL , 40 μL , and 60 μL . The same buffer volume was added to the assay solution instead of the tested amounts in the blank solution. Galantamine hydrobromide was used as a positive control during the assay. To start the reactions, ≈ 0.03 U/mL of AChE (electric eel) was transferred into the reaction mixture. The reaction was monitored for 2 minutes using a spectrophotometer (Cary 60 single beam spectrophotometer, Agilent Technologies, USA) at 412 wavelengths. Following the activity obtained from the blank, as the percentage of the reaction rate, the enzymatic activity was calculated. The data obtained from the linear section of the initial 60 s were used to calculate the activities. The subtraction of the ratio of the sample activity versus blank activity from 100 calculated the AChE inhibitory activity. The results of the experiments were given as mean \pm standard deviation of three parallel experiments. The analysis was performed only on methanolic extract due to the solubility and turbidity problem of the dichloromethane extract.

RESULTS and DISCUSSION

GC-MS analysis of essential oil

The essential oil composition of the plant species was detected to be rich in saturated fatty acids. The major compound of *Isatis floribunda* was determined as dodecanoic acid represents about 28.6%. Nanocosane, hexadecenoic acid, and tetradecanoic acid were also detected at about 11.0%, 10.0%, and 8.4%, respectively. In addition, methyl octadecenoate, decanoic acid, and hexahydrofarnesyl acetone were detected at the ratio of 4.8%, 4.6%, and 3.5%, respectively. Due to insufficient essential oil, a bioactivity assay could not be performed. Therefore, an extract of the plant material was obtained for further analysis. GC-MS chromatogram is presented in "Figure 1".

Table 1. Compounds determined by GC-MS of *Isatis floribunda* essential oil, major compounds were presented in bold font type

No	¹ RT	² RRI	³ RRI Lit	Compound	%	⁴ Identification method
1	17,879	1229	1232	(E)-2 Hexanal	0.12	RI, MS
2	25,232	1403	1400	Nonanal	0.06	RI, MS
3	26,674	1443	1441	(E)-2-Octenal	0.09	RI, MS
4	32,831	1627	1624	cis-Dihydrocarvone	0.07	RI, MS
5	33,23	1640	1638	β -Cyclocitral	0.09	RI, MS
6	33,445	1647	1645	cis-Isodihydrocarvone	0.05	RI, MS
7	34,01	1666	1664	Phenylacetaldehyde	0.29	RI, MS
8	34,319	1676	1671	(Z)- β -Farnesene	0.08	RI, MS
9	35,397	1712	1709	α -Terpineol	0.15	RI, MS
10	37,019	1769	1765	Napthalene	0.46	RI, MS
11	38,236	1813	1815	Methyl dodecanoate	0.06	RI, MS
12	39,255	1850	1845	Anethole	0.08	RI, MS
13	39,325	1853	1853	Ethyl dodecanoate	0.13	RI, MS
14	39,798	1871	1868	(E)-Geranyl acetone	0.75	RI, MS
15	42,17	1962	1958	(E)- β -Ionone	0.59	RI, MS
16	42,544	1976	1956	2-Methyl-5-(1,1,5-trimethyl-5-hexenyl)furan	0.26	RI, MS
17	43,572	2017	1968	β -Ionol	0.09	RI, MS
18	43,778	2025		3,4-Dehydro- β -ionone	0.34	MS
19	44,496	2055	2048	3,4-Dimethoxystyrene	1.15	RI, MS
20	44,605	2059	2057	Ethyl tetradecanoate	0.07	RI, MS
21	44,92	2072	2084	Octanoic acid	0.11	RI, MS
22	45,619	2101	2096	(E)-Methyl cinnamate	0.06	RI, MS
23	45,717	2105	2100	Heneicosane	0.48	Ac, RI, MS
24	46,5	2138	2131	Hexahydrofarnesyl acetone	3.50	RI, MS
25	47,451	2179	2179	1-Tetradecanol	0.06	RI, MS
26	47,553	2183	2170	3,4-Dimethyl-5-pentylidene-2(5H)-furanone	0.52	RI, MS
27	47,668	2188		Megastigmatrienone isomer*	0.07	MS
28	47,913	2199	2198	1-Docosene	0.09	RI, MS
29	48,506	2225	2218	p-Vinylguaicol	0.09	RI, MS
30	48,666	2232		Megastigmatrienone isomer*	0.46	MS

Table 1. Compounds determined by GC-MS of *Isatis floribunda* essential oil, major compounds were presented in bold font type (continued)

31	48,754	2236	2241	Heptadecanal	0.24	RI, MS
32	49,421	2266	2262	Ethyl hexadecanoate	0.11	RI, MS
33	49,863	2286	2296	Decanoic acid	4.69	RI, MS
34	50,314	2305	2300	Tricosane	0.50	Ac, RI, MS
35	50,996	2334		Megastigmatrienone isomer*	0.22	MS
36	51,567	2358	2353	Octadecanal	0.06	RI, MS
37	52,365	2391	2384	Farnesyl acetone	0.59	RI, MS
38	52,757	2407	2400	Tetracosane	0.34	Ac, RI, MS
39	53,866	2446	2431	Methyl octadecanoate	4.84	RI, MS
40	54,19	2458	2467	Ethyl octadecanoate	0.37	RI, MS
41	55,392	2500	2503	Dodecanoic acid	28.62	RI, MS
42	55,601	2506	2500	Pentacosane	3.08	RI, MS
43	57,808	2571	2560	Ethyl 3-hydroxytridecanoate	0.30	RI, MS
44	58,127	2580	2583	Methyl linolenate	0.07	RI, MS
45	59,14	2608	2600	Hexacosane	0.17	Ac, RI, MS
46	59,357	2613	2613	Ethyl linolenate	0.09	RI, MS
47	59,814	2624	2622	Phytol	0.43	RI, MS
48	61,284	2659	2670	Ethyl eicosanoate	1.51	RI, MS
49	61,521	2665	2670	Tetradecanoic acid	0.06	RI, MS
50	63,429	2709	2700	Heptacosane	2.62	Ac, RI, MS
51	63,557	2712	2713	Tetradecanoic acid	8.54	RI, MS
52	68,141	2809	2800	Octacosane	0.41	Ac, RI, MS
53	68,581	2818	2822	Pentadecanoic acid	0.13	Ac, RI, MS
54	70,485	2856	2857	Palmito- γ -lactone	0.25	RI, MS
55	73,307	2911	2900	Nonacosane	11.03	Ac, RI, MS
56	74,049	2925	2931	Hexadecanoic acid	9.99	RI, MS
57	78,749	3013	2290	Docosanol	0.52	RI, MS

RT: Retention Time, RRI: Relative Retention Index according to n-alkanes, RRI Lit: Relative Retention Index according to the literature. Identification Method; MS: According to mass similarity, RI: According to the similarity of RRI with the literature, Ac: Co-injection of the authentic compound

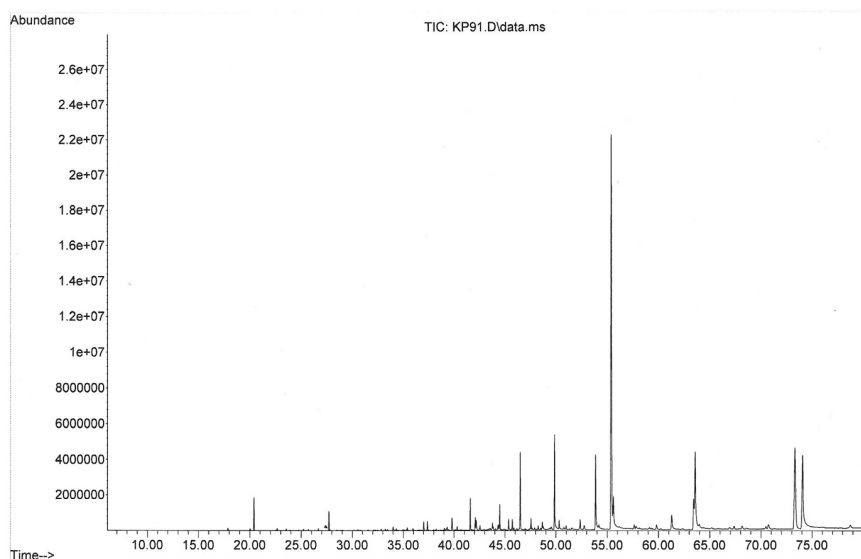


Figure 1. GC-MS chromatogram of *Isatis floribunda* essential oil

***In vitro* acetylcholinesterase inhibition assay**

Due to the solubility and turbidity problem of the dichloromethane extract, an AChE inhibitory assay was performed only on the methanolic extract. All the tested volumes showed inhibitory activity. 20 μ L, 40 μ L, and 60 μ L volumes demonstrated 35.17%, 50.44%, and 57.323% inhibition, respectively. The IC_{50} of positive control, galantamine hydrobromide, was calculated with the same equation which was 2.01 μ L. The detailed results of the inhibitory assay are demonstrated in “Table 2”.

Table 2. AChE inhibition results of the methanolic extracts of *Isatis floribunda*

Tested Volume, μ L	[Inh], mg/mL	Inhibition, %	Standard Deviation \pm
0	0	0	0
20	0.083	35.17	0.39
40	0.166	50.44	0.208
60	0.25	57.323	1.258

IC_{50} was calculated as 0.16 mg/mL according to the equation obtained from the AChE inhibitory curve. The AChE inhibition curve of *Isatis floribunda* is demonstrated in “Figure 2”.

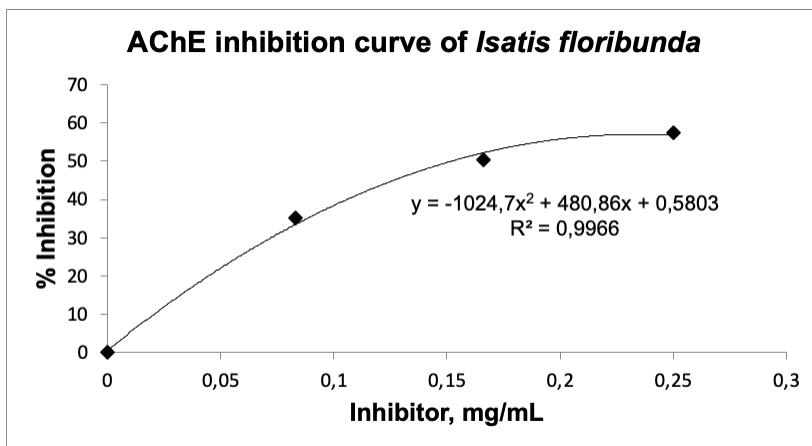


Figure 2. AChE inhibition curve of *Isatis floribunda* methanolic extract

Alzheimer's is the most common neurodegenerative disease and has become a major health issue worldwide. However, there is still no cure for the disease, which makes it one of the top research topics. AChE inhibition is one of the crucial treatment options in treating Alzheimer's disease. However, the use of the current AChE inhibitors is limited due to the side effects¹³. Physostigmine has a narrow therapeutic index and shorter half-life and has numerous side effects, including diarrhea, abdominal cramps, increased sweating, and increased saliva production. Due to these reasons, its use in treating Alzheimer's disease is not approved anymore¹⁴. Tacrine also has a short half-life and has side effects of nausea, vomiting, clumsiness, and diarrhea. Also, blood monitoring is required during Tacrine treatment because of its hepatotoxic effects. Therefore, its use in Alzheimer's disease is not continued anymore^{15,16}. Another AChE inhibitor is Donepezil, which demonstrates side effects of insomnia, loss of appetite, muscle cramps, and muscle weakness. Additionally, it has been detected that those patients under Donepezil treatment have experienced low blood pressure, severe vomiting, breathing problems, and bradycardia. Therefore, its use in Alzheimer's disease is not approved in some countries. Rivastigmine also has similar side effects, including stomach pain, weight loss, diarrhea, loss of appetite, nausea, and vomiting¹³. Lastly, Galantamine is shown to be effective in treating cognitive symptoms of Alzheimer's disease. However, it also has side effects that limit medication use including convulsions, irregular breathing, stomach cramps, watery eyes, and confusion¹⁷. Due to all these reasons, novel AChE inhibitors are needed to treat Alzheimer's disease.

Natural sources such as plant species have been used to treat several diseases for centuries. As such, extracts, essential oils, and fractions of different plant species, including the families of Acanthaceae, Amaranthaceae, and Amaryllidaceae have been used for their AChE inhibitory effect¹⁸. As previously mentioned, Rivastigmine and Galantamine are also medications derived from natural sources¹⁹. In previous studies, *Isatis floribunda* extracts have been tested for its antioxidative, antibacterial, and cytotoxic properties⁴. However, the AChE inhibitory effect of the plant species is still a mystery. Therefore, *Isatis floribunda* extracts have been tested against their AChE inhibitory effect in this study. It has been found that the methanolic extracts have a promising AChE inhibitory effect with the IC₅₀ value of 0.16 mg/mL according to the equation obtained from the AChE inhibitory curve. Previous studies have shown that the chemical composition of methanolic extracts of *Isatis floribunda* contains catechin, chlorogenic acid, caffeic acid, quercetin, and p-coumaric acid⁴. Therefore, it has been believed that the AChE inhibition of *Isatis floribunda* extract is due to these compounds²⁰⁻²³. Additionally, the essential oil composition of *Isatis floribunda* was demonstrated to be rich in fatty acids, which are also potent AChE inhibitors^{24,25}. Therefore, sufficient *Isatis floribunda* essential oil could also demonstrate an AChE inhibitory effect.

This study determined the AChE inhibitory effect of *Isatis floribunda* extracts. Methanolic extracts of the plant species have demonstrated strong AChE inhibition with the IC₅₀ of 0.16 mg/mL, which shows that it is a promising plant material for developing new AChE inhibitors. Additionally, it has been shown that the essential oil composition of *Isatis floribunda* is rich in compounds which are potentially effective AChE inhibitors. Therefore, even though the essential oil yield was insufficient, it is believed that the essential oil of the plant species could be a promising AChE inhibitor in the future. In conclusion, *Isatis floribunda* is a promising plant species in developing novel AChE inhibitors in the treatment of Alzheimer's disease.

STATEMENT OF ETHICS

Ethics approval is not required in this study, as no human and experimental animal samples are not involved.

CONFLICT OF INTEREST STATEMENT

Declared none.

AUTHORS CONTRIBUTIONS

Concept: E.G., Y.Y., K.P., Design: E.G., Y.Y., K.P., Data Collection and Processing: E.G., Y.Y., M.A., K.P., Analysis or Interpretation: E.G., Y.Y., K.P., Literature Search: E.G., Writing: E.G.

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REFERENCES

1. Nair RR, Kalariya T, Chanda S. Antibacterial activity of some selected Indian medicinal flora. *Turk J Biol*, 2005;29(1):41-47.
2. Dar MI, Khan FA, Rehman F, Masoodi A, Ansari AA, Varshney D, Naushin F, Naikoo MI. Roles of Brassicaceae in phytoremediation of metals and metalloids. *Phytoremediation*, 2015(1):201-215. Doi: 10.1007/978-3-319-10395-2_14
3. Soundararajan P, Kim J. Anti-carcinogenic glucosinolates in cruciferous vegetables and their antagonistic effects on prevention of cancers. *Molecules*, 2018;23(11):2983. Doi: 10.3390/molecules23112983
4. Karakoca K, Asan Ozusaglam M, Cakmak YS, Karaman Erkul S. Antioxidative, antimicrobial and cytotoxic properties of *Isatis floribunda* Boiss. ex Bornm. extracts. *EXCLI J*, 2013;(12):150-167.
5. Radwan HM, Shams KhA, Tawfik WA, Soliman AM. Investigation of the glucosinolates and lipids constituents of *Cakile maritima* (Scope) growing in Egypt and their biological activity. *Res J Med Med Sci*, 2008;3(2):182-187.
6. Cavdar H, Senturk M, Guney M, Durdagi S, Kayik G, Supuran CT, Ekinici D. Inhibition of acetylcholinesterase and butyrylcholinesterase with uracil derivatives: kinetic and computational studies. *J Enzyme Inhib Med Chem*, 2019;34(1):429-437. Doi: 10.1080/14756366.2018.1543288
7. Adewusi EA, Moodley N, Steenkamp V. Antioxidant and acetylcholinesterase inhibitory activity of selected Southern African medicinal plants. *South Afr J Bot*, 2011;77(3):638-644. Doi: 10.1016/j.sajb.2010.12.009.
8. Fu AL, Li Q, Dong ZH, Huang SJ, Wang YX, Sun MJ. Alternative therapy of Alzheimer's disease via supplementation with choline acetyltransferase. *Neurosci Lett*, 2004;368(3):258-262. Doi: 10.1016/j.neulet.2004.05.116
9. Dave KR, Syal AR, Katyare SS. Tissue Cholinesterases. A comparative study of their kinetic properties. *Z Für Naturforschung C*, 2000;55(1-2):100-108. Doi: 10.1515/znc-2000-1-219
10. Diken ME, Yilmaz B. Inhibitory effect on acetylcholinesterase and toxicity analysis of some medicinal plants. *Int J Second Metab*, 2022;9(1):27-42. Doi: 10.21448/ijsm.1032863
11. Colovic MB, Krstic DZ, Lazarevic-Pasti TD, Bondzic AM, Vasic VM. Acetylcholinesterase inhibitors: pharmacology and toxicology. *Curr Neuropharmacol*, 2013;11(3):315-335. Doi: 10.2174/1570159X11311030006
12. Ellman GL, Courtney KD, Andres V, Featherstone RM. A new and rapid colorimetric determination of acetylcholinesterase activity. *Biochem Pharmacol*, 1961;7(2):88-95. Doi: 10.1016/0006-2952(61)90145-9
13. Sharma K. Cholinesterase inhibitors as Alzheimer's therapeutics (Review). *Mol Med Rep*, 2019;20(2):1479-1487. Doi: 10.3892/mmr.2019.10374
14. Thal LJ, Fuld PA, Masur DM, Sharpless NS. Oral physostigmine and lecithin improve memory in Alzheimer disease. *Ann Neurol*, 1983;13(5):491-496. Doi: 10.1002/ana.410130504
15. Farlow M, Gracon SI, Hershey LA, Lewis KW, Sadowsky CH, Dolan-Ureno J. A controlled trial of tacrine in Alzheimer's disease. *JAMA*, 1992;268(18):2523-2529. Doi: 10.1001/jama.1992.03490180055026
16. Watkins PB. Hepatotoxic effects of tacrine administration in patients with Alzheimer's disease. *JAMA*, 1994;271(13):992. Doi: 10.1001/jama.1994.03510370044030

17. Mehta M, Adem A, Sabbagh M. New acetylcholinesterase inhibitors for Alzheimer's disease. *Int J Alzheimers Dis*, 2012;2012:1-8. Doi: 10.1155/2012/728983
18. Murray A, Faraoni M, Castro M, Alza N, Cavallaro V. Natural AChE inhibitors from plants and their contribution to Alzheimer's disease therapy. *Curr Neuropharmacol*, 2013;11(4):388-413. Doi: 10.2174/2F1570159X11311040004
19. Devi P, Syad AN. Botanic: a potential source of new therapies for Alzheimer's disease?. *Bot Targets Ther*, 2014;2014(4):11-26. Doi: 10.2147/BTAT.S33554
20. Oboh G, Agunloye OM, Akinyemi AJ, Ademiluyi AO, Adefegha SA. Comparative study on the inhibitory effect of caffeic and chlorogenic acids on key enzymes linked to Alzheimer's disease and some pro-oxidant induced oxidative stress in rats' brain-*in vitro*. *Neurochem Res*, 2013;38(2):413-419. Doi: 10.1007/s11064-012-0935-6
21. Okello EJ, Mather J. Comparative kinetics of acetyl- and butyryl-cholinesterase inhibition by green tea catechins |relevance to the symptomatic treatment of Alzheimer's disease. *Nutrients*, 2020;12(4):1090. Doi: 10.3390/nu12041090
22. Orhan IE. Cholinesterase inhibitory potential of quercetin towards Alzheimer's disease - a promising natural molecule or fashion of the day? - a narrowed review. *Curr Neuropharmacol*, 2021;19(12):2205-2213. Doi: 10.2174/2F1570159X1866620119153807
23. Ouattara N, Tiero Meda RN, Hilou A, Guenne S, Konate K, Coulibaly AY, Kiendrebeogo M, Millogo JF, Nacoulma OG. Anti-acetylcholinesterase and antioxidant activities and HPLC-MS analysis of polyphenol from extracts of *Nelsonia canescens* (Lam.) Spreng. *Asian Pac J Trop Di*, 2013;3(5):382-388. Doi: 10.1016%2FS2222-1808(13)60088-2
24. Akay MB, Şener K, Sari S, Bodur E. Inhibitory action of omega-3 and omega-6 fatty acids alpha-linolenic, arachidonic and linoleic acid on human erythrocyte acetylcholinesterase. *Protein J*, 2023;42(2):96-103. Doi: 10.1007/s10930-022-10088-z
25. Itriago Yanes CV, Melo Filho AA, Ribeiro PRE, Reis Melo ACG, Takahashi JA, Ferraz VP, Mozombite DMS, Santos RC. Inhibition of acetylcholinesterase and fatty acid composition in *Theobroma grandiflorum* seeds. *Orbital: Electron J Chem*, 2017;9(3):127-130. Doi: 10.17807/orbital.voio.894